Effects of Amphiphile Topology on Aggregation **Properties:** Distinctive Behavior of Contrafacial Amphiphiles

David G. Barrett and Samuel H. Gellman*

S. M. McElvain Laboratory of Organic Chemistry Department of Chemistry, University of Wisconsin 1101 University Avenue, Madison, Wisconsin 53706

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Amphiphilic molecules play important roles in many settings.¹ Biological systems, for example, use lipid-based bilayers for structural and functional definition of living cells and subcellular compartments. Synthetic surfactants find applications in many personal care products and industrial processes. The most important properties of amphiphiles, which stem from their aggregation and interaction with other molecules, are determined to a significant extent by the topological relationship between hydrophilic and lipophilic surfaces.^{1,2} Amphiphiles with the most common topology, involving a compact polar "headgroup" and a flexible nonpolar "tail", aggregate cooperatively in aqueous solution to form micelles or vesicles. Alternative amphiphile topologies can lead to distinctive and useful properties. For example, the cholic acids (steroids with rigid polar and nonpolar surfaces) are physiologically important in the uptake and excretion of insoluble lipids³ and have proven to be unique tools for the manipulation and purification of proteins.^{4,5} Cholate itself aggregates less cooperatively and with greater size polydispersity than typical compact headgroup/flexible tail amphiphiles;6 this nonmicellar mode of association is undoubtedly related to the cholic acids' functional merits.

The exploration of new amphiphilic architectures is a matter of continuing interest and importance.7 We have been examining synthetic amphiphiles that have rigid planar structures with one polar face and one nonpolar face,8 species that we designate "contrafacial amphiphiles".9 (Cholate and derivatives do not completely fit this description, because the carboxylate is attached at the end of a flexible alkyl appendage that can extent approximately laterally from the steroid nucleus.) Here we report the aggregation properties of two synthetic contrafacial amphiphiles, 1a and 2a, and of a related structure bearing a short

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(5) The synthesis of glycosylated derivatives of cholic acid and allocholic acid, with enhanced hydrophilicity on one side, has been reported: (a) Kahne, D.; Walker, S.; Cheng, Y.; Van Engen, D. J. Am. Chem. Soc. 1989, 111, 6881. (b) Cheng, Y.; Ho, D. M.; Gottlieb, C. R.; Kahne, D.; Bruck, M. A. J. Am. Chem. Soc. 1992, 114, 7319.

(6) (a) Roda, A.; Hofmann, A.F.; Mysels, K. J. J. Biol. Chem. 1983, 258, 6362. (b) Mukerjee, P.; Moroi, Y.; Murata, M.; Yang, A. Y. S. Hepatology 1984, 4, 61S.

(7) (a) Regen et al. have developed an imaginative membrane-directed drug design strategy based upon the "molecular harpoon" concept; for leading references, see: Naka, K.; Sadownik, A.; Regen, S. L. J. Am. Chem. Soc. 1993, 115, 2278. (b) Careful choice of polar group allows redox-switched vesicle formation: Muñoz, S.; Gokel, G. W. J. Am. Chem. Soc. 1993, 115, 4899 and references therein. (c) "Gemini surfactants": Menger, F. M.; Littau, A. J. Am. Chem. Soc. 1991, 113, 1451. (d) "Bolaamphiphiles": Fuhrhop, J.-H.; Fritsch, D. Acc. Chem. Res. 1986, 19, 130.

(8) Stein, T. M.; Gellman, S. H. J. Am. Chem. Soc. 1992, 114, 3943.
(9) Kahne et al. (ref 5b) have used the term "facial amphiphile" to describe cholic acids and their derivatives. For molecules like 1a and 2a, we prefer 'contrafacial amphiphile", a term that emphasizes the fact that all polar functionality is facially disposed.

flexible tail (3a).¹⁰ The behavior of 1a is compared to that of isomer 4a, which is planar and rigid but has its polar group peripherally rather than facially disposed. The topologies of contrafacial amphiphile 1a and isomer 4a are indicated schematically below.



Aggregation of 1a-4a in D₂O could be detected by monitoring ¹H NMR signals as a function of amphiphile concentration. In each case, ¹H chemical shifts were independent of concentration in dilute solutions (<5 mM), but most or all resonances of each amphiphile began to move upfield at higher concentrations, signaling the onset of intermolecular interactions.¹¹ For each amphiphile, the changes in chemical shift with increasing concentration were gradual, indicating that the associations were not highly cooperative, i.e., that these aggregation processes do not correspond to the classical models of micelle formation.¹ This noncooperative behavior is consistent with observations reported for rigid amphiphiles of more conventional topology, including benzoate and benzenesulfonate derivatives.¹² According to the ¹H NMR data, the onset of association occurs in the range 20-40 mM for isomers 1a and 4a and in the range 5-10 mM for 2a and 38.11

Aggregation of 1a-4a was probed also by monitoring uptake of the water-insoluble azo dye orange OT as a function of amphiphile concentration (Figure 1).13 Aggregates of 1a did not solubilize dye even at the limit of this contrafacial amphiphile's solubility (0.54 M). In contrast, aggregates of 4a began to solubilize dye at ca. 0.33 M. The onset of dye uptake for 3a was ca. 0.06 M, but no dye solubilization was detected for 2a up to 0.16 M, which is near this amphiphile's solubility limit.¹⁴ The difference in behavior between isomers 1a and 4a demonstrates that the contrafacial amphiphile topology does indeed confer unique properties: the structure of the aggregates formed by 1a must differ fundamentally from the structure of the aggregates formed by 4a, with only the latter capable of providing a microenvironment that can solubilize the dye. Comparison of 2a and 3a shows that the attachment of a short flexible segment to a rigid contrafacial amphiphile unit can exert a significant effect on aggregation and solubilization properties.

For each of the amphiphiles we have studied, aggregation is detected by 1H NMR at concentrations at which dye solubilization

(14) For aqueous solutions of 3a, the following dye amphiphile ratios were observed: 80 mM 3a, 1:7700; 166 mM 3a, 1:520. For aqueous solutions of 4a, the ratios were the following: 425 mM 4a, 1:24 000; 525 mM 4a, 1:6200.

⁽¹⁾ For leading references, see: Myers, D. Surfactant Science and Technology; 2nd ed.; VCH Publishers, Inc.: New York, 1992. (b) Fendler, J. H. Membrane Mimetic Chemistry; John Wiley & Sons: New York, 1982. (2) (a) Mukerjee, P. J. Pharm. Sci. 1974, 63, 972. (b) Israelachvili, J. N.;

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⁽¹⁰⁾ Compound 1b was prepared as previously described: Barrett, D. G.; Liang, G.-B.; Gellman, S. H. J. Am. Chem. Soc. 1992, 114, 6915. (For the original synthesis of 1b, see: Vogel, E. Spec. Publ.—Chem. Soc. 1967, 21, 113.) The syntheses of acids 2b and 3b were similar to the previous route, except that intermediate allyl rather than methyl esters were used.

⁽¹¹⁾ Data may be found in the supplementary material.

⁽¹²⁾ For leading references on the association of benzene carboxylates and sulfonates, see: (a) Saleh, A. M.; El-Khordagui, L. K. Int. J. Pharm. 1985, 24, 231. (b) Balasubramanian, D.; Srinivas, V.; Gaikar, V. G.; Sharma, M. M. J. Phys. Chem. 1989, 93, 3865. (c) Pyrene sulfonate in aqueous solution: Menger, F. M.; Whitesell, L. G. J. Org. Chem. 1987, 52, 3793. (13) Schott, H. J. Phys. Chem. 1966, 70, 2966 and references therein.



Figure 1. Orange OT solubilization as a function of amphiphile concentration in aqueous solution: 1a (O), 2a (Δ), 3a (+) and 4a (×). The lines for 1a and 2a are results of linear regression; the lines for 3a and 4a are arbitrary. For each ampt⁻¹-hile, aqueous solutions with varying amphiphile concentrations and excess suspended orange OT were agitated gently at room temperature for 2-3 days. The solutions were then filtered through cotton to remove undissolved dye, aliquots of each filtrate were diluted with 4 vol of absolute EtOH, and the absorbance was measured at 500 nm. Orange OT has an absorbance maximum near this wavelength, but amphiphiles 1a-4a do not. The electronic absorption bands of bridged annulenes 1a-3a (but not naphthalene derivative 4a) "tail" weakly out to 500 nm, which causes the small amount of absorbance seen for concentrations of 1a and 2a. The inset shows an expanded plot at low concentrations for 2a and 3a.

does not occur. This behavior differs from that of typical micelleforming amphiphiles, for which critical micelle concentration (CMC) values determined by these two methods are usually comparable.¹ Our observations suggest that aggregates of 3aand 4a must grow to a minimum size before a dye-solubilizing microenvironment is created, behavior that is consistent with the noncritical nature of the aggregation processes for these amphiphiles.

The crystal structures of acid forms **1b**-4b were determined to allow comparison of the molecular packing of the methanobridged annulene and naphthalene skeletons. Neighboring naphthyl units in 4b display a "herringbone" arrangement, as is observed for naphthalene itself¹⁵ (interplanar angle for 4b = 48°; for naphthalene = 51°). Figure 2 shows that such a "herringbone" juxtaposition occurs as well in crystalline 1b. Similar aromatic-



Figure 2. Ball-and-stick representation of a neighboring pair of molecules in the crystal lattice of 1b. The angle between the mean planes of the annulene rings is 67° .

aromatic juxtapositions are observed in crystalline 2b and 3b.¹⁶ In light of the qualitative difference in solution behavior between 4a and bridged annulene-based contrafacial amphiphiles 1a and 2a, it is interesting that the faces and edges of bridged annulenes behave similarly to the analogous surfaces of more conventional aromatic species in the solid state. The nature of the intermolecular juxtapositions in solution aggregates of 1b-4b is unclear. It has been assumed that aromatic sulfonates and carboxylates engage in parallel "stacking" interactions in aqueous aggregates, $1^{2a,b}$ but there appear to be no structural data to support this assumption.

We have shown that the contrafacial amphiphile topology confers unique solution properties relative to more traditional amphiphile topologies. The fact that **1a** and **2a** present substantial expanses of nonpolar surface to their aqueous surroundings but, in their aggregated states, do not create a microenvironment that can solubilize a hydrophobic moiety suggests that these and related molecules may display interesting nondisruptive behavior toward proteins and biological membranes.

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Supplementary Material Available: Crystallographic details for 1b-4b, and plots of ¹H NMR chemical shifts vs concentration for aqueous solutions of 1a-4a (34 pages); listing of observed and calculated structure factors for 1b-4b (20 pages). Ordering information is given on any current masthead page.

^{(15) (}a) Abrahams, S. C.; Monteath Robertson, J.; White, J. B. Acta Crystallogr. 1949, 2, 238. (b) Brock, C. P.; Dunitz, J. D. Acta Crystallogr. 1982, B38, 2218 and references therein. (c) For a general discussion of aromatic hydrocarbon crystal packing, see: Desiraju, G. R.; Gavezzotti, A. Acta Crystallogr. 1989, B45, 473.

⁽¹⁶⁾ Barrett, D. G.; Desper, J. M.; Hayashi, R. K., unpublished results.